Coverslip Preparation for Neuronal Culture

Monday

1. Place coverslips in ceramic racks (plastic drawer at electrophoresis area).
2. Rinse with RO water in Pyrex tray (TC room cabinet).
3. Blot off excess water and transfer into dish containing 70% Nitric Acid (in the fume hood).
4. Leave Overnight.

* Ensure that coverslips are fully immersed in 70% Nitric Acid.

Tuesday

1. Rinse coverslips 4X with RO water (by transferring each rack of coverslips into fresh RO water tray each time).

2. Sterilize each coverslip by dipping in Absolute EtOH and letting them dry in the laminar flow hood.
   - Lay out the coverslips on C-fold tissue towels in Pyrex Trays.

3. Once dried, UV Sterilize for at least 20mins.

4. Coat the coverslips with PolyLysine (1mg/ml; stock in TC room fridge) overnight.
   - Place sterilized coverslips (7 coverslips per 100mm Bacterial Petri Dish)
   - Add between 200ul-400ul of PolyLysine on top of each coverslip (enough to cover the entire surface
     Of each coverslip).
   - Leave the coated coverslips in the Petri Dishes on the table outside the laminar Flow hood.

Wednesday

1. In the laminar flow hood, remove the polylysine and wash the coverslips 2X with sterile RO water for 2hr each (by adding on top of each coverslip).
2. Add 10ml of neuronal plating medium and leave in incubator till ready to use.